

FLUID-BASED BIOMARKERS FOR NEURODEGENERATIVE DISEASES



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INTRODUCTION

The prevalence of neurodegenerative diseases is expected to rise alongside the increase in life expectancy in most of the world. According to a publication from The Alzheimer's Disease International, the estimated global prevalence of dementia was 36 million in 2010 which rose to 43.8 million by 2016 and is estimated increase to 130 million by 2050.¹ The most common neurodegenerative disease is Alzheimer's disease (AD), accounting for 60-70% of dementia cases.^{2,3} This is followed by vascular dementias which account for up to 20% of dementia patients.³ Less common causes of dementia include frontotemporal dementias (FTD) and dementia with Lewy Bodies (LBD). The second most common neurodegenerative disease is Parkinson's disease (PD), with an estimated global prevalence of over six million.²

The diagnosis and management of patients with dementia presents challenges as the underlying cause of disease can clinically present in a manner that masks its true nature or reflect influence of confounding comorbidities.³ Moreover, misdiagnosis

can lead to suboptimal care and treatment. Therefore, it is important to have reliable biomarkers that increase the accuracy of clinical diagnosis. Additionally, accurate and timely diagnosis of neurodegenerative diseases in the clinical setting provides the ability to optimize treatment strategies and allows patients the opportunity to join appropriate clinical treatment trials.

It is becoming increasingly common for patients to seek earlier medical attention with the onset of mild cognitive impairment (MCI) - characterized by impairment in a single cognitive domain (usually memory), or moderate impairment in several cognitive domains, but without functional impairment sufficient to meet the criteria for dementia.⁴ Nearly 30 -50% of patients presenting with MCI have incipient AD, however the underlying etiology is difficult to determine without biomarkers.² In regard to PD, while the accuracy of a clinical diagnosis is around 80%, correctly identifying other parkinsonian disorders is more difficult.^{2,5,6} Therefore, specific biomarkers for neurodegenerative indications are needed to improve the diagnostic workup of these diseases and allow for the treatment effect monitoring and repeated longitudinal tracking of disease progression over the course of clinical trials. The following biomarker sections detail current fluid-based biomarkers for various neurodegenerative disease pathologies.

CURRENT BLOOD AND CSF-BASED BIOMARKERS FOR NEURODEGENERATIVE DISEASES

Biomarkers of Amyloid Beta Pathology

Amyloid beta (A β) plaques formed within the extracellular space of telencephalic structures are one of the major neuropathological features in AD.⁷ Studies using positron emission tomography (PET) have shown that A β accumulation starts ~ 20 years before the onset of AD dementia.^{7,8}

Different A β species can be reliably measured in cerebrospinal fluid, including A β 40 and A β 42. Before A β -PET shows abnormal findings, CSF levels of A β 42 are decreased (~50%) in AD.⁹ Using A β 42 to A β 40 ratios (A β 42/ A β 40) results in high concordance with



A β -PET (primarily above 90%).¹⁰ Moreover, CSF A β 42 measurements when used in conjunction with A β 40 or phosphorylated tau, (P-tau), can be used to predict the development of AD dementia in patients with mild cognitive impairment with high accuracy.¹¹

Current clinical routine A β measurement procedures are based on enzyme linked immunosorbent assays (ELISA) or immunoassays on other technology platforms. Recently, reference materials based on human CSF that are certified for the mass concentration of A β 42 became available and are used to ensure equivalence of results across methods and platforms.¹² Additionally, an international consensus protocol for the collection and handling of CSF has been established to avoid false abnormal values due to confounding preanalytical factors.¹³

However, while A β biomarkers in CSF are already in clinical use, the collection procedure (lumbar puncture) is invasive and is not widely available outside of specialist centers. Therefore, considerable effort has been made to develop blood-based assays for measuring cerebral A β pathologies.² As the levels of these markers in plasma/serum are expectedly lower than those found in CSF, ultra-sensitive detection methodologies are required for quantification in this matrix. In cerebral A β pathology, plasma levels of A β 42/ A β 40 are decreased by only 10–20% while CSF levels are decreased by 40–60%.^{14–16} This is most likely due to the production of A β outside of the brain that may affect plasma levels. As such, CSF A β 42/ A β 40 ratios show higher diagnostic accuracy as compared to plasma levels. However, combinations of plasma A β 42/A β 40 with other plasma CNS markers such as P-tau and GFAP, have shown strong associations with cerebral A β pathology.^{17,18}

Biomarkers of Tau Pathology

In addition to A β accumulation, neurofibrillary tangles of tau protein are well-known neuropathologic characteristic of AD. Several different soluble species of tau can be quantified in CSF.² Some of these include total tau, tau phosphorylated at threonine 181 (p-tau181), and tau phosphorylated at threonine 217. While total tau (T-tau) concentration is increased in AD patients, T-tau is not AD specific as it is also found markedly increased in Creutzfeldt-Jacobs disease (CJD) and temporarily increased with brain trauma or stroke.¹⁹

On the other hand, p-tau (p-tau181 and p-tau217) has been shown to be selectively increased in AD 20 and correlated with the formation of neurofibrillary tangles in the brain.^{21,22} When combined with CSF A β 42, CSF levels of T-tau and p-tau181 increase the diagnostic validity for AD with a combined sensitivity and specificity of 85–90%²³ and predict the cognitive decline in patients with MCI.²⁴ Specifically, CSF p-tau181 has been found to be notably useful at differentiating AD from other dementias, including Lewy body disease (LBD), frontotemporal dementia (FTD), and vascular dementia.²⁵ Recently, it has been shown that CSF p-tau217 displays a moderately stronger association with tau-tangle load and disease severity and may distinguish AD dementia from other dementias with even higher accuracy than P-tau181.^{26,27}

While tau biomarkers are commonly assessed in CSF, ultrasensitive assays that can reliably quantify T-tau and p-tau in blood have recently been developed. In addition to the minimally invasive collection process for plasma/serum samples (as opposed to CSF), clinic-based cohort studies have shown that plasma p-tau (181 and 217) can differentiate AD dementia from other neurodegenerative diseases with high accuracy.² Plasma p-tau217 performed alongside CSF AD biomarkers and tau-PET imaging exhibits a diagnostic accuracy of around 90%,²⁸ while increased plasma p-tau181 levels have been associated with ensuing cognitive decline and conversion to AD dementia in patients with MCI.^{29,30} Moreover, longitudinal studies have demonstrated that plasma p-tau217 increases during early AD and continues to increase over time in both preclinical and prodromal stages of AD, indicating its potential use in the objective monitoring of AD progression.³¹ Finally, plasma levels of P-tau181 and P-tau217 were shown to be associated with the amount of A β plaques (early AD stages) and tau-tangles (later AD stages) in recent neuropathology-based and PET-based studies.³² Additionally, plasma p-Tau and p-Tau/ A β 42 ratios have been used to predict cognitive and executive function decline in levodopa treated Parkinson's disease patients.³³



Biomarkers of Alpha Synuclein Pathology

CSF oligomeric α -synuclein is found to be consistently higher in PD patients when compared with healthy controls.³⁵ Additionally, several studies have found that phosphorylated α -synuclein is increased in CSF from PD patients when compared to healthy controls. However, the diagnostic accuracy for these α -synuclein species when used alone was found to be unsatisfactory for potential use in clinical practice.³⁴ An increase in diagnostic accuracy was observed when combined with other neurodegenerative markers (ratio of oligomeric α -synuclein to total α -synuclein, together with phosphorylated α -synuclein and phosphorylated p-tau).³⁶ Due to the very low concentrations of phosphorylated α -synuclein in CSF, mass spectrometry remains one of the primary methods for quantifying this biomarker.³⁴

Seeding assays such as Protein-Misfolding Cyclic Amplification (PMCA) and Real-Time Quaking Induced Conversion (RT-QuIC) that have been developed to indirectly reveal misfolded prion protein and other prion-like proteins – can be used to detect pathogenic α -synuclein aggregates in CSF.^{2,34}

Recent studies have evaluated the clinical use of quantifying α -synuclein in peripheral blood samples from PD patients using an ultrasensitive single molecule array technology (SIMOA by Quanterix)³⁷ and an immunomagnetic reduction (IMR) assay,³⁸ with both studies concluding that peripheral blood α -synuclein levels may differentiate between healthy controls and patients with PD. However, given the considerable overlap with controls and other neurodegenerative diseases, the use of α -synuclein alone as a single biomarker is not presently sufficient as a diagnostic marker in clinical practice and trials and therefore must be used in conjunction with other documented biomarkers.³⁴

Biomarkers of Neurodegeneration

Several biofluid markers for neurodegeneration have come to light. Of note, neurofilaments have been studied extensively in neurodegenerative indications and are useful markers of acute and chronic neuronal injury.^{39,40} Neurofilament light (NfL) and Phosphorylated neurofilament heavy chain (pNFH) proteins have been extensively studied as diagnostic and prognostic indicators of ALS. The protein pNFH is released in the CSF and subsequently into blood as

a result of axonal damage and has been found to be advantageous over other biomarkers for ALS disease identification due to its high specificity for neuronal cell damage.⁴⁰

Neurofilament light chain (NfL) can also be measured in CSF and blood and reflects axonal degradation, regardless of cause - with elevated levels observed in ALS, FTD, and atypical parkinsonian disorders.^{2,40} Additionally, NfL levels are also elevated in AD with studies demonstrating that NfL levels in CSF and serum correlate with one another and are increased during presymptomatic stages of autosomal dominant AD.⁴¹ Notably, NfL can serve as an indicator of the intensity of ongoing neurodegeneration as increased levels of NfL are associated with faster disease progression and higher brain atrophy rates in most neurodegenerative indications.^{41,42} Recent studies have supported a role for plasma and CSF NfL levels as a treatment response biomarker for multiple sclerosis and spinal muscular atrophy, respectively, with reductions associated with the clinical effectiveness of the drug.^{43,44}

CSF levels of the postsynaptic neurogranin protein serve as a potential biomarker of neurodegenerative disease as this analyte has been shown to be selectively increased in AD patients and can predict future cognitive decline.⁴⁵⁻⁴⁷ Synaptosomal associated protein (SNAP25) has also been labeled as a potential biomarker of synaptic dysfunction and neurodegeneration. Increased CSF levels of SNAP25 were found in AD patients versus healthy controls⁴⁸ and may predict progression to AD in patients with MCI.⁴⁹ Finally, CSF Visinin-like protein-1 (VILIP-1) is also found elevated in AD patients as compared to healthy controls^{50,51} and has demonstrated potential utility as a marker of neuronal injury. CSF VILIP-1 and the ratio to A β 42 (VILIP-1/A β 42) were found to predict rates of global cognitive decline similarly to tau and the tau/A β 42 ratio.⁵²

Biomarkers of Oxidative Stress

Oxidative stress due to increased levels of reactive oxygen and nitrogen species, together with compromised antioxidant defense mechanisms, is prominent in many neurodegenerative diseases⁵³. Elevated concentrations of MDA and NME have been reported in the plasma and cerebrospinal fluid (CSF) of PD patients⁵⁴ and increased levels of 8-OHdG and nitrotyrosine have been observed in the serum and leukocytes of HD patients.^{53,55,56}



Additionally, about 20% of familial ALS patients carry mutations in the superoxide dismutase (SOD) gene, resulting in the production of ROS.^{53,57} The overall level of oxidative damage can be evaluated by monitoring the oxidative stress marker, MDA, in blood and CSF using the TBARS (thiobarbituric acid-reacting substances) assay.⁵³

DJ-1 is a multifunctional protein associated with several cellular processes. Loss of function PARK7 gene mutations, the gene that encodes DJ-1, are linked to familial forms of PD and lead to oxidative stress.^{2,53} As such, plasma levels of DJ-1 have been evaluated in PD patients in addition to other oxidative stress-related biomarkers such as uric acid.⁵⁸ Urate is a strong antioxidant that may exert protective activity towards the development of PD as elevated levels of serum uric acid conferred a lower risk of developing PD.⁵⁸⁻⁶⁰

Biomarkers of Microglia Activation, Neuroinflammation, and Blood-Brain-Barrier Dysfunction

Neuroinflammation is a pathological feature of a wide range of central nervous system (CNS) diseases including neurodegenerative diseases.⁶¹ Microglia and astrocytes are primary cellular drivers and regulators of neuroinflammation. Different blood and CSF molecules have been proposed as reliable markers for glial cell activation within CNS. These include markers of reactive astrocytes (glial fibrillary acidic protein (GFAP)), microglia activation (soluble triggering receptor expressed on myeloid cell 2 (sTREM2)), microglial mobilization (YKL-40, chitinase-3-like protein 1), and astrocytic activation (monocyte chemoattractant protein 1 (MCP-1)).^{2,61}

Plasma levels of GFAP were found to be increased in FTD caused by progranulin mutations⁶² and individuals with A β pathology.¹⁷ Moreover, plasma GFAP levels served as a predictive factor for future cognitive decline and conversion to AD dementia in cognitively unimpaired individuals.⁶³ Additionally, it has been shown that microglia are activated during the preclinical stage of AD as indicated by increased levels of CSF sTREM2, with increased levels of MCP-1 and YKL-40 observed in the later phase of the disease.⁶⁴ Finally, neuroinflammation panels capable of providing multiplex analysis of cytokines and chemokines in CSF are available on multiplexing platforms offered by Mesoscale Discovery (MSD®) and Luminex®.



Disruption of blood-brain barrier integrity is observed in several neurodegenerative diseases. This disruption is largely observed through specific MRI techniques that depend on the leakage of gadolinium-based contrast reagents into the brain.⁶⁵ However, measurement of fluid biomarkers has also been utilized. The ratio of CSF albumin to plasma albumin has been shown to be elevated in non-AD dementia disorders and used to demonstrate the permeability of the blood-brain barrier.⁶⁶ More recently, CSF levels of soluble PDGFR β (sPDGFR β) has been proposed as a marker of capillary pericyte damage and BBB leakiness, with elevated levels associated with cognitive decline and AD pathology.⁶⁷

Genetic Biomarkers

AD is classified into two subtypes based on the age of disease onset: early age onset which accounts for approximately 1-5% of AD cases (EOAD, before the age of 65 years) and late onset (LOAD, after the age of 65 years).⁵⁸ EOAD is frequently caused by mutations in the amyloid precursor protein (APP), Presenilin 1 (PSEN1), or Presenilin 2 (PSEN2).⁶⁸ In the majority of LOAD cases, AD is assumed to be developed due to a combination of genetic variants contributing to



disease formation, in addition to other factors such as environmental exposure, lifestyle, and aging. Currently, the E4 allele of the apolipoprotein E gene (APOE) is the most characterized risk factor for LOAD.⁵⁸ Specifically, of the three APOE alleles (E2, E3, and E4) one copy of the E4 allele increases the risk of AD by four-fold while two copies increase the risk by 12-fold.⁶⁹ APOE mutation detection and genotyping can be routinely performed in central and reference laboratories on DNA extracted from EDTA-anticoagulated whole blood via sanger sequencing.

In PD, approximately 15% of patients have a family history of the disorder caused by mutations in the following genes: LRRK2, SNCA, PARK2, PARK7, GBA, or PINK1. More recently, RNA-based biomarkers (miRNAs) have been found as potential markers of PD. These include the miRNAs, miR-7 and miR-153, which were found to modulate α -synuclein mRNA.⁵⁸

CONCLUSION

Over the past two decades considerable advancement has been made regarding fluid-based biomarker research in the field of neurodegeneration. As clinical trials of potential disease-modifying therapeutics targeting neurodegenerative disorders have shifted focus towards the preclinical and early stages of these diseases, accessibility to sensitive and specific biomarkers that could be used for initial screening phases within a multistage diagnostic process for disease differentiation, or could aid in earlier diagnosis, subject selection, and disease tracking has become of paramount importance.⁷²

Most progress has been made with CSF biomarkers, with lumbar puncture considered to be a safe and generally well-tolerated procedure, and an international consensus protocol for the collection and handling of CSF has been established.¹³ However, CSF use may be limited by contraindications such as the use of anticoagulants, subject non-compliance, and the availability of this procedure at sites outside of specialist centers.^{72,73} As such, measurement of CNS biomarkers in more accessible biofluids, such as serum and plasma, would provide a less invasive means for sample material collection and would facilitate access to repeated longitudinal samples for tracking disease progression.⁷² Additionally, collection of serum/plasma is possible for sites outside of specialist centers.

As a result of the selectivity of the BBB which prevents the free passage of molecules between the CNS and blood compartments and affects the blood to CSF volume ratio,⁵⁰ the concentration of CNS-derived proteins in blood is much lower than that found in CSF. This concentration difference has made it difficult to detect CNS proteins in blood samples using standard ELISA assays. Fortunately, in recent years several ultrasensitive immunoassays with superior analytical sensitivity have been developed, such as the automated single molecule array (SIMOA) by Quanterix, which has allowed for the reliable detection of CNS biomarkers in peripheral blood samples (biomedicines paper). At the present time the following analytes A β 40, A β 42, BDNF, GFAP, NfL, α -synuclein, P-tau 231, P-tau 181, total tau and SNAP-25 can be measured by the SIMOA technology in peripheral blood samples. Recently, the ultrasensitive SIMOA technology was utilized by Eli Lilly to measure p-tau 217 using their proprietary antibodies and a licensing agreement was made to provide Quanterix access to Lilly's P-tau 217 antibody technology.⁷⁴ Medpace Reference Laboratories can support these biomarker assays and help further the clinical development of disease-modifying neurodegenerative disease pharmaceuticals.

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Medpace provides customized, high quality central laboratory services to pharmaceutical and biotech clinical development industries. Our four wholly owned laboratories offer full-service support to seven continents for phase I-IV studies. We have extensive experience from small and simple clinical trials to those that are large, global, and complex. Our wholly owned global laboratory facilities, standardized testing platforms, comprehensive test menu, and stellar project management teams allow Medpace to set up fully customized projects for our clients. Combined with Medpace Clinical Research Organization expertise, we provide a fully integrated solution for your clinical development needs.



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With laboratories in the US, Europe, China and Singapore, Medpace Central Laboratories has the global reach and capabilities to conduct CNS studies in concert with Medpace CRO or as standalone service.

Our test menu includes validated biomarkers associated with prevalent CNS disorders: A β 40, A β 42, GFAP, NfL, and P-tau 181. These markers

are analyzed on the Quanterix HD-X instrument using the ultra-sensitive, automated, single molecule array (SIMOA) methodology that allows for accurate and precise detection in serum, plasma, and cerebral spinal fluid (CSF). Fully validated 10-plex Proinflammatory (IFN- γ , IL-1 β , IL-2, IL-4, IL-6, IL-8, IL-10, IL-12p70, IL-13, and TNF- α), 9-plex cytokine (GM-CSF, IL-1 α , IL-5, IL-7, IL-12/23p40, IL-15, IL-17A, TNF- β , and VEGF-A), and 7-plex chemokine (IP-10, MCP-1, MCP-4, MDC, MIP-1 α , MIP-1 β , and TARC) panels analyzed via the Mesoscale Discovery platform are also on our test menu in addition to CNS markers YKL-40, α 2-macroglobulin, and APOE genotyping (via Sanger sequencing). In addition, Medpace Central Laboratories can validate new biomarkers in a variety of matrices (serum, plasma, CSF, PBMCs, etc.) quickly and efficiently – typically with an industry leading timeline of 12-14 weeks.

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REFERENCES

1. Prince, M. et al. World Alzheimer Report 2015. The Global Impact of Dementia: An Analysis of Prevalence, Incidence, Cost and Trends. <https://www.alzint.org/u/WorldAlzheimerReport2015.pdf> (Alzheimer's Disease International, 2015).
2. Hansson, O. Biomarkers for neurodegenerative diseases. *Nat Med.* 27, 954-963 (2021).
3. Hunter, C. A. et al. Medical costs of Alzheimer's disease misdiagnosis among US Medicare beneficiaries. *Alzheimers Dement.* 11, 887-895 (2015).
4. Petersen, R. C. Clinical practice. Mild cognitive impairment. *N. Engl. J. Med.* 364, 2227-2234 (2011).
5. Rizzo, G. et al. Accuracy of clinical diagnosis of Parkinson disease: a systematic review and meta-analysis. *Neurology* 86, 566-576 (2016).
6. 6Respondek, G. et al. Validation of the Movement Disorder Society criteria for the diagnosis of 4-repeat tauopathies. *Mov. Disord.* 35, 171-176 (2020).
7. Villemagne, V. L. et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *Lancet Neurol.* 12, 357-367 (2013).
8. Gordon, B. A. et al. Spatial patterns of neuroimaging biomarker change in individuals from families with autosomal dominant Alzheimer's disease: a longitudinal study. *Lancet Neurol.* 17, 241-250 (2018).
9. Olsson, B. et al. CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol.* 15, 673-684 (2016).
10. Hansson, O. et al. Advantages and disadvantages of the use of the CSF Amyloid β (A β) 42/40 ratio in the diagnosis of Alzheimer's disease. *Alzheimers Res. Ther.* 11, 34 (2019).
11. Mattsson, N. et al. Clinical validity of cerebrospinal fluid A β 42, tau, and phospho-tau as biomarkers for Alzheimer's disease in the context of a structured 5-phase development framework. *Neurobiol. Aging* 52, 196-213 (2017).
12. Boulo, S. et al. First amyloid β 1-42 certified reference material for re-calibrating commercial immunoassays. *Alzheimers Dement.* 11, 1493-1503 (2020).
13. Hansson, O. et al. The Alzheimer's Association international guidelines for handling of cerebrospinal fluid for routine clinical measurements of amyloid β and tau. *Alzheimers Dement.* <https://doi.org/10.1002/alz.12316> (2021).
14. Schindler, S. E. et al. High-precision plasma β -amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* 93, e1647-e1659 (2019).
15. Nakamura, A. et al. High-performance plasma amyloid- β biomarkers for Alzheimer's disease. *Nature* 554, 249-254 (2018). (58)
16. Palmqvist, S. et al. Performance of fully automated plasma assays as screening tests for Alzheimer disease-related β -amyloid status. *JAMA Neurol.* 76, 1060-1069 (2019).
17. Verberk, I. M. W. et al. Combination of plasma amyloid beta(1-42/1-40) and glial fibrillary acidic protein strongly associates with cerebral amyloid pathology. *Alzheimers Res. Ther.* 12, 118 (2020).
18. Janelidze, S. et al. Detecting amyloid positivity in early Alzheimer's disease using combinations of plasma A β 42/A β 40 and P-tau. *Alzheimers Dement.* 18, 283-293 (2022).



-
19. 1Gabelli, C. Blood and cerebrospinal fluid biomarkers for Alzheimer's disease. *J Lab Precis Med* [online], 5 (2020).
 20. Blennow, K. & Zetterberg, H. Biomarkers for Alzheimer's disease: current status and prospects for the future. *J. Intern. Med.* 284, 643–663 (2018).
 21. Buerger, K. et al. CSF phosphorylated tau protein correlates with neocortical neurofibrillary pathology in Alzheimer's disease. *Brain.* 11, 3035-3041 (2006).
 22. de Souza, LC. et al. CSF tau markers are correlated with hippocampal volume in Alzheimer's disease. *Neurobiol Aging.* 33, 1253-1257 (2012).
 23. Blennow, K. et al. Amyloid biomarkers in Alzheimer's disease. *Trends Pharmacol Sci.* 36, 297-309 (2015).
 24. Blennow, K. et al. Predicting clinical decline and conversion to Alzheimer's disease or dementia using novel Elecsys A β (1–42), pTau and tTau CSF immunoassays. *Sci. Rep.* 9, 19024 (2019).
 25. Kang, JH. Et al. Clinical utility and analytical challenges in measurement of cerebrospinal fluid amyloid-beta(1-42) and tau proteins as Alzheimer disease biomarkers. *Clin Chem.* 59, 903-916 (2013).
 26. Janelidze, S. et al. Cerebrospinal fluid p-tau217 performs better than p-tau181 as a biomarker of Alzheimer's disease. *Nat. Commun.* 11, 1683 (2020).
 27. Hanes, J. et al. Evaluation of a novel immunoassay to detect p-tau Thr217 in the CSF to distinguish Alzheimer disease from other dementias. *Neurology* 95, e3026–e3035 (2020).
 28. Palmqvist, S. et al. Discriminative accuracy of plasma phospho-tau217 for Alzheimer disease vs other neurodegenerative disorders. *JAMA* 324, 772–781 (2020).
 29. Janelidze, S. et al. Plasma P-tau181 in Alzheimer's disease: relationship to other biomarkers, differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. *Nat. Med.* 26, 379–386 (2020).
 30. Karikari, T. K. et al. Diagnostic performance and prediction of clinical progression of plasma phospho-tau181 in the Alzheimer's Disease Neuroimaging Initiative. *Mol. Psychiatry* 26, 429–442 (2021).
 31. Mattsson-Carlsson, N. et al. Longitudinal plasma p-tau217 is increased in early stages of Alzheimer's disease. *Brain* 143, 3234–3241 (2020).
 32. Mattsson-Carlsson, N. et al. Soluble P-tau217 reflects amyloid and tau pathology and mediates the association of amyloid with tau. *EMBO Mol. Med.* <https://doi.org/10.15252/emmm.202114022> (2021).
 33. Liu, C. et al. CSF tau and tau/Ab42 predict cognitive decline in Parkinson's disease. *Parkinsonism Relat Disord.* 21, 271-276 (2015).
 34. Parnetti, L. et al. CSF and blood biomarkers for Parkinson's disease. *Lancet Neurol.* 18, 573-586 (2019).
 35. Eusebi, P. et al. Diagnostic utility of cerebrospinal fluid α -synuclein in Parkinson's disease: a systematic review and meta-analysis. *Mov Disord.* 32,1389-1400 (2017).
 36. Majbour, NK. et al. Oligomeric and phosphorylated alpha-synuclein as potential CSF biomarkers for Parkinson's disease. *Mol Neurodegener.* 11, 7 (2016).
 37. Ng, ASL. et al. Plasma alpha-synuclein detected by single molecule array is increased in PD. *Ann Clin Transl Neurol.* 15, 615-619 (2019).



-
38. Chang, CW. et al. Plasma and Serum Alpha-Synuclein as a Biomarker of Diagnosis in Patients with Parkinson's Disease. *Front Neurol.* 10, 1388 (2020).
 39. Bacioglu, M. et al. Neurofilament light chain in blood and CSF as marker of disease progression in mouse models and in neurodegenerative diseases. *Neuron* 91, 56-66 (2016).
 40. Poesen, K and Van Damme, P. Diagnostic and Prognostic Performance of Neurofilaments in ALS. *Front Neurol.* 9, 1167 (2019).
 41. Preische, O. et al. Serum neurofilament dynamics predicts neurodegeneration and clinical progression in presymptomatic Alzheimer's disease. *Nat. Med.* 25, 277-283 (2019).
 42. Khalil, M. et al. Neurofilaments as biomarkers in neurological disorders. *Nat. Rev. Neurol.* 14, 577-589 (2018).
 43. Delcoigne, B. et al. Blood neurofilament light levels segregate treatment effects in multiple sclerosis. *Neurology* 94, e1201-e1212 (2020).
 44. Olsson, B. et al. NFL is a marker of treatment response in children with SMA treated with nusinersen. *J. Neurol.* 266, 2129-2136 (2019).
 45. Portelius, E. et al. Cerebrospinal fluid neurogranin concentration in neurodegeneration: relation to clinical phenotypes and neuropathology. *Acta Neuropathol.* 136, 363-376 (2018).
 46. Portelius, E. et al. Cerebrospinal fluid neurogranin concentration in neurodegeneration: relation to clinical phenotypes and neuropathology. *Acta Neuropathol.* 136, 363-376 (2018).
 47. Lista, S. et al. Cerebrospinal Fluid Neurogranin as a Biomarker of Neurodegenerative Diseases: A Cross-Sectional Study. *J Alzheimers Dis.* 59, 1327 -1334 (2017).
 48. Brinkmalm, A. et al. SNAP-25 is a promising novel cerebrospinal fluid biomarker for synapse degeneration in Alzheimer's disease. *Mol Neurodegener.* 9,53 (2014).
 49. Zou, K. et al. Current Biomarkers for Alzheimer's Disease: From CSF to Blood. *J Pers Med.* 10, 85 (2020).
 50. Koníčková, D. et al. Biomarkers of Neurodegenerative Diseases: Biology, Taxonomy, Clinical Relevance, and Current Research Status. *Biomedicines.* 10, 1760 (2022).
 51. Lee, JM. et al. The brain injury biomarker VLP-1 is increased in the cerebrospinal fluid of Alzheimer disease patients. *Clin. Chem.* 54, 1617 -1623 (2008).
 52. Tarawneh, R. et al. CSF VILIP-1 predicts rates of cognitive decline in early Alzheimer disease. *Neurology.* 78, 709-719 (2012).
 53. Sidorova, Y and Domanskyi, A. Detecting Oxidative Stress Biomarkers in Neurodegenerative Disease Models and Patients. *Methods Protoc.* 3, 66 (2020).
 54. Selley, ML. (E)-4-hydroxy-2-nonenal may be involved in the pathogenesis of Parkinson's disease. *Free Radic Biol Med.* 25, 169-174 (1998).
 55. Browne, SE. et al Oxidative stress in Huntington's disease. *Brain Pathol.* 9, 147-163 (1999).
 56. Chen, C. et al. Increased oxidative damage and mitochondrial abnormalities in the peripheral blood of Huntington's disease patients. *Biochem. Biophys. Res. Commun.* 359, 335-340 (2007).
 57. Rosen, DR. et al. Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature.* 362, 59-62 (1993).



-
58. Jeromin, A and Bowser, R. Biomarkers in Neurodegenerative Diseases. *Adv Neurobiol.* 15, 491-528 (2017).
59. Shen, C. et al. Serum urate and the risk of Parkinson's disease: results from a meta-analysis. *Can J Neurol Sci.* 88, 73–79 (2012).
60. de Lau, LM. et al. Serum uric acid levels and the risk of Parkinson disease. *Ann Neurol.* 58, 797–800 (2005).
61. Gaetani, L. et al. CSF and Blood Biomarkers in Neuroinflammatory and Neurodegenerative Diseases: Implications for Treatment. *Trends Pharmacol Sci.* 41, 1023-1037 (2020).
62. Heller, C. et al. Plasma glial fibrillary acidic protein is raised in progranulin-associated frontotemporal dementia. *J. Neurol. Neurosurg. Psychiatry* 91, 263–270 (2020).
63. Verberk, I. M. W. et al. Serum markers glial fibrillary acidic protein and neurofilament light for prognosis and monitoring in cognitively normal older people: a prospective memory clinic-based cohort study. *Lancet Healthy Longevity* 2, E87–E95 (2021).
64. Nordengen, K. et al. Glial activation and inflammation along the Alzheimer's disease continuum. *J Neuroinflammation.* 16, 46 (2019).
65. Elschot, E. P. et al. A comprehensive view on MRI techniques for imaging blood–brain barrier integrity. *Invest. Radio.* 56, 10–19 (2021).
66. Janelidze, S. et al. Increased blood-brain barrier permeability is associated with dementia and diabetes but not amyloid pathology or APOE genotype. *Neurobiol. Aging* 51, 104–112 (2017).
67. Nation, D. A. et al. Blood–brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat. Med.* 25, 270–276 (2019).
68. Goate, A. et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 349, 704–706 (1991).
69. Strittmatter, W.J. et al. Apolipoprotein E: high avidity binding to B-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 90, 1977–1981 (1993).
70. Khoo, S.K. et al. Plasma-based circulating microRNA biomarkers for Parkinson's disease. *J Parkinsons Dis* 2, 321–331 (2012).
71. Mouradian, M.M. MicroRNAs in Parkinson's disease. *Neurobiol Dis.* 46, 279–284 (2012).
72. Obrocki, P. et al. Perspectives in fluid biomarkers in neurodegeneration from the 2019 biomarkers in neurodegenerative diseases course—a joint PhD student course at University College London and University of Gothenburg. *Alzheimers Res Ther.* 12, 20 (2020).
73. Duits, FH. et al. Performance and complications of lumbar puncture in memory clinics: Results of the multicenter lumbar puncture feasibility study. *Alzheimers Dement.* 12, 154-163 (2016).
74. Romine, P. Press release: Quanterix Announces New Agreements With Lilly To Advance Alzheimer's Disease Diagnosis And Treatment. PAN Communications. <https://www.quanterix.com/press-releases/quanterix-announces-new-agreements-with-lilly-to-advance-alzheimers-disease-diagnosis-and-treatment/> (2022).

